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NO. 1, 2022

# Science Could Bring Sustainability to Your Table

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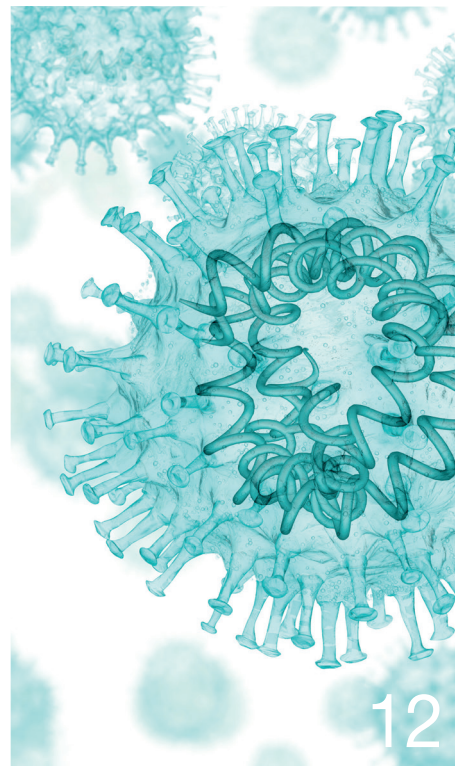
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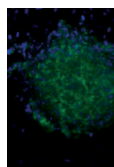


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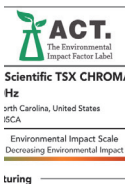
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# A Brighter Future for Farming and Renewable Energy

By Gina Wynn

Climate change, globalization, and a generation of farmers nearing retirement age are just some of the trends that have driven many farms out of business in recent years. But the increasing use of agrivoltaics is at least one development that more and more farmers view as trending in their favor. The technology could light the way for a new era of renewable energy use, improved water conservation, and the creation of a more sustainable food system.

Agrivoltaics involves growing agricultural crops underneath photovoltaic solar installations. It's a type of low-impact solar development, among others, that the U.S. Department of Energy's National Renewable Energy Lab (NREL) has been studying over the past 10 years. Through its Innovative Site Preparation and Impact Reductions on the Environment (InSPIRE) project, NREL has been investigating the economic and ecological impact of dual land use practices, including pairing solar installations with agricultural crops, native vegetation growth, and pollinator habitats, according to [nrel.gov](http://nrel.gov).

## A Model for Small Farms

One of about a dozen NREL agrivoltaic sites in the U.S., Jack's Solar Garden in Boulder County, Colorado, has installed rows of 3,200 solar panels on a four-acre field. Around 40 types of plants, including tomatoes, garlic, lettuce, radishes, peppers, beets, and kale grow beneath the panels that are mounted on six- to eight-foot-high posts spaced far enough apart for a tractor to navigate between them.

Former Peace Corps volunteer and U.S. Agency for International Development Natural Resources Officer Byron Kominek owns the 24-acre family farm that had been producing alfalfa and hay for nearly fifty years. When these crops had stopped turning a profit, Kominek saw agrivoltaics as a means of supporting his family while helping the environment. He worked with NREL and researchers from Colorado State University (CSU) to build "a model for other small farms that want to keep their soils productive while taking advantage of the economic benefits that clean energy production can provide," according to the *Solar Power World* article, "Largest agrivoltaic research project in U.S. advances renewable energy while empowering local farmers" by HansenRE.

## Worth the Challenge

Switching to agrivoltaics wasn't easy, however. CSU and NREL helped Kominek work with county regulators to change the designation of his land so that he would have approval to install

the solar array. Then, to finance the \$2 million panels, Kominek had to put his farm and the array up for collateral. But so far, his investment has been paying off, according to the *npr.org* article "This Colorado 'solar garden' is literally a farm under solar panels."

Kominek and the researchers found that many of the plants under the panels were thriving because of the intermittent shade the structures provided. That shade also helped reduce evaporation of the irrigation water. The water that did evaporate helped cool the baking solar panels and made them work more efficiently.

Because other agrivoltaic farms have recorded the same reduction of water use, western states that have been battling drought have become particularly interested in agrivoltaics. A University of Arizona study showed that some crops grown beneath solar panels needed 50 percent less water. Using less water benefits the environment and also cuts irrigation expenses.

## A Cleaner, Greener Energy Source

As for economics, Kominek sells 1.2 megawatts of power back to the local grid, enough to power 300 homes for a year. The ability to generate and sell power provides farmers like Kominek a stable, additional source of income in an unpredictable agriculture industry. In addition, by using solar power to create energy rather than fossil fuels, Kominek is contributing to meeting the country's renewable energy targets.

Now with close to two billion dollars allocated for renewable power through the Bipartisan Infrastructure Deal, the U.S. is aiming to expand renewable energy use, including solar. A recent Oregon State University (OSU) study found that farms could generate 20 percent of U.S. electricity if 1 percent or 13,000 square miles of agricultural land (an area about the size of Maryland) were converted to agrivoltaics, according to [today.oregonstate.edu/news](http://today.oregonstate.edu/news).

Also, non-agricultural land for solar development is becoming scarce. By 2030, utility-scale solar could cover almost 2 million acres of land in the U.S., according to NREL.

The advantages of agrivoltaics are clear to researchers. In addition to providing a practical alternative to monopolizing land solely for energy creation or food production, it offers a ray of hope for small to mid-sized farms looking to grow profits.

*Gina Wynn is a Thermo Fisher Scientific staff writer.*



# Optical Clearing for Improved Confocal Imaging of Thick Specimens

Developed almost 70 years ago, confocal microscopy has become a mainstay for imaging fluorescently labeled biological samples. Widefield microscopy illuminates biological samples with a diffuse field of light that excites fluorophores above and below the focal plane of interest, resulting in a blurry, less resolved image. By placing a pinhole in a conjugate focal plane between the sample and the light detector (camera or PMT), confocal microscopy blocks contaminating out-of-focus light, thus improving axial (z) resolution (Figure 1). This improved resolution means that finer z-sections can be taken deeper within thick samples, such as spheroids and resected tissue, and three-dimensional biological relationships can be better elucidated.

Poor image quality and resolution (e.g. signal-to-noise ratio) can result not only from excited out-of-focus fluorophores but also from biological matter such as lipids and proteins. In the latter case, in-focus light diffracts as it travels through the sample, reducing the amount of signal that is detected. This diffractive effect is quantified by the refractive index, which is a measure of how the speed of light is influenced as it passes through a material. Matching the refractive index of the mounting medium and the sample with the microscope optics as close as possible will aid in minimizing diffracted light, which in turn will improve the quality of the image.

Many clearing agents and mounting media have been developed to help reduce diffracted light by altering the refractive index of the sample so that they better match the refractive index of the microscope optics. Selecting the most appropriate clearing agent requires consideration of multiple factors, including sample thickness, imaging depth, fluorophore type, and vessel format and material. Some clearing agents function by simply exchanging the primarily water-based intracellular environment with a liquid that has a better-matched refractive index, such as glycerol. Alternatively, some clearing agents improve sample clarity by extracting lipids (delipidation). Clearing agents that simply exchange the intracellular aqueous environment are minimally invasive and tend to favor smaller samples and those where fluorescent proteins are used. While clearing agents that delipidate may improve imaging depth to a greater degree, these agents typically employ harsh detergents and electrophoretic devices to draw lipids away from the sample. Some delipidating clearing agents use organic solvents, which may be incompatible with fluorescent protein-based samples. Additionally, organic solvent-based clearing agents may not be compatible with imaging vessels that use glass alternatives as the imaging window, such as cycloolefin. This technical note describes the use of a refractive index-correcting clearing agent (Figure 2) to improve imaging depth of thick samples such as HT-1080 spheroids.

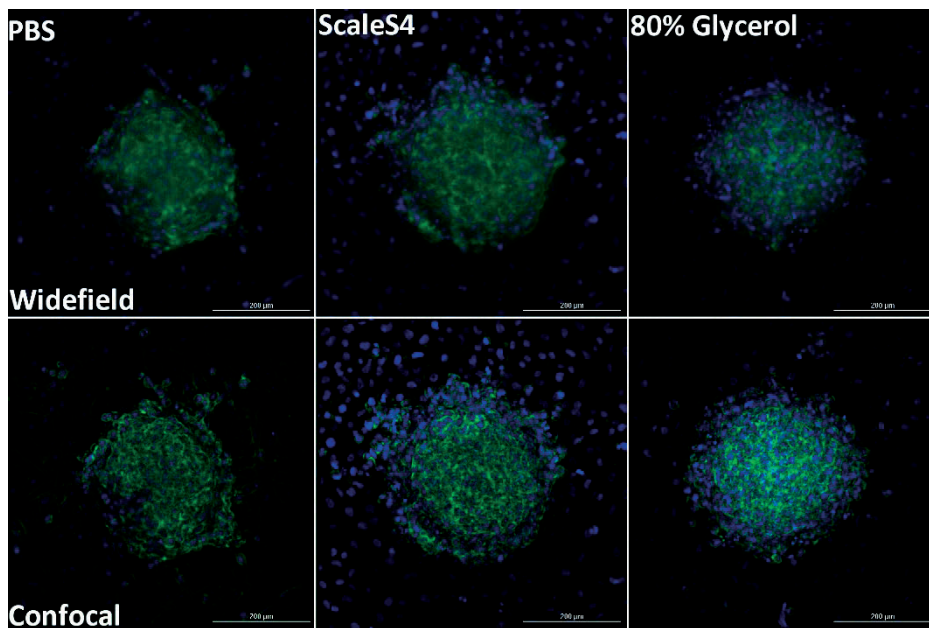


Figure 1. Adhered HT-1080 spheroids. Adhered 1000-cell HT-1080 spheroids were fixed and stained with Hoechst 33342 and Alexa Fluor 488-phalloidin, then buffer (PBS) was either maintained or exchanged for ScaleS4 clearing agent or 80% glycerol. Z-stacks were then captured at 20x (0.45NA) in either widefield or confocal (60  $\mu$ m disc) mode. Maximum intensity projections of the entire z-stack were then generated.

## Sample Preparation for Imaging HT1080 Spheroids

HT-1080 spheroids were formed by seeding 1,000 cells into ultra-low-



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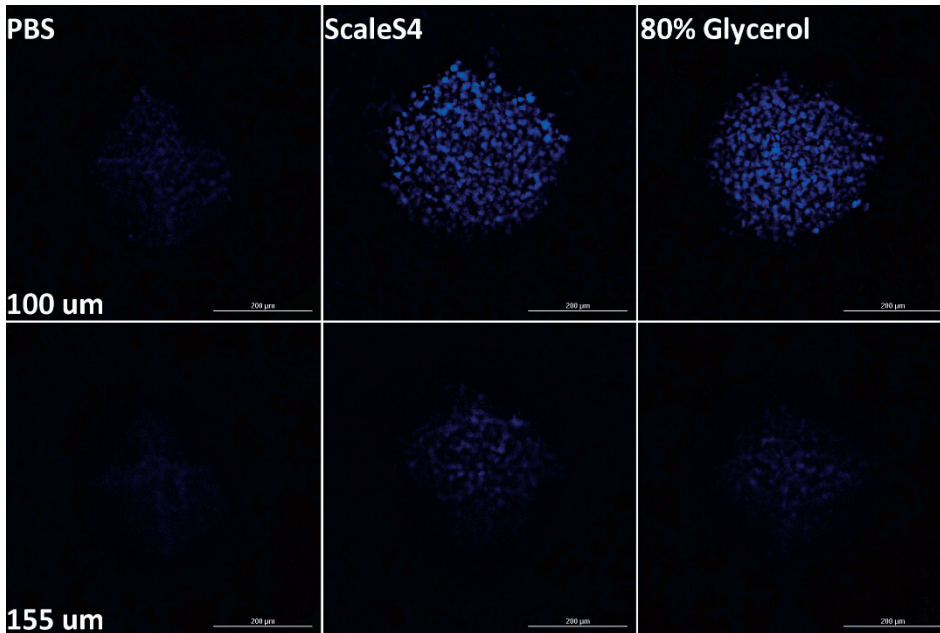


Figure 2. Imaging depth limits of adhered HT-1080 spheroids in different imaging media. Z-slices from each adhered spheroid in Figure 1 were selected when nuclei become unresolvable for PBS (upper panels), or for ScaleS4 or 80% glycerol (lower panels).

attachment (ULA) U-bottom 96-well plates, #4520, from Corning and allowed to coalesce for 24 hours in Gibco Advanced DMEM from Thermo Fisher Scientific at 37°C. Spheroids were then transferred to clear, flat-bottom 96-well plates, #204626, from Agilent Technologies (1 spheroid per well) and allowed to settle and establish adhesion at RT for one hour. Plates containing spheroids were then transferred back to the 37°C incubator and allowed to attach to the culture area and spread overnight. Attached spheroids with migrating cell regions were fixed with 4% PFA for 30 minutes, then permeabilized with 0.5% for one hour. To achieve full penetration of dyes, spheroids were stained with Hoechst 34580 and Alexa Fluor 488-phalloidin

overnight at 4°C. Spheroids were then washed, and PBS was either maintained or exchanged for either a homemade mounting medium (80% glycerol, 20 mM tris, pH 8.0, 0.5% n-propyl gallate, 0.05% NaN<sub>3</sub>) or the ScaleS4 (40% D-sorbitol, 10% glycerol, 4M urea, 15% DMSO, 0.5% n-propyl gallate, 0.05% NaN<sub>3</sub>) (Figure 1).

## Imaging Procedure and Processing

Confocal and widefield z-stack images of attached HT-1080 spheroids were captured at 40x using the Cytation C10 confocal imaging reader and Gen5 software in Manual Mode (IMM). The dynamic range of each channel was

maximized by selecting the “show saturated pixels” button. The exposure settings for each channel were adjusted such that they were within range, and there were no saturated pixels. The z-range was determined by toggling below and above the sample until the structure of interest (in this example, a spheroid) faded in intensity. The z-step size was set using Nyquist sampling recommendations; for imaging Alexa Fluor 488 using the 40x air objective (NA = 0.6), steps sizes were determined to be 1 μm for widefield and 0.6 μm for confocal. Images were then subjected to five iterations of deconvolution using a point spread function based on the objective. Deconvolved images were then pre-processed to reduce background signal. The rolling ball size for background reduction for Hoechst 34580 (DAPI channel) and Alexa Fluor 488-phalloidin was set to 25 and 50 μm, respectively. Equivalent z-heights for all three conditions (PBS, 80% glycerol, ScaleS4) was compared to determine imaging depth limits (Figure 2).

Confocal microscopy is a powerful mode of imaging that improves image quality by removing out-of-focus light. However, sources of diffracted fluorescent light inherent to the sample prevent this improvement from being truly realized. Use of clearing agents allows matching the refractive index of the sample, in this case HT-1080 spheroids, with the refractive index of the optical system allowing imaging depth and resolution to be improved.



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### Reference

1. Recommended by Germany's Federal Institute for Risk Assessment.

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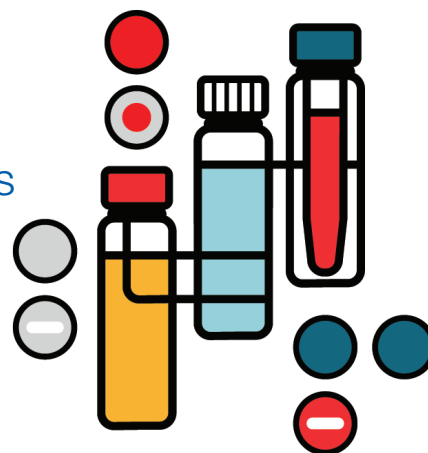
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# How the ACT Label Supports Your Sustainability Goals

For many scientists who see themselves as environmentally conscious, the scale of their total carbon footprint can be a wake-up call. Laboratories are under increasing pressure to adopt more sustainable practices, lower their environmental impact, and contribute to a cleaner and safer society. But what can be done to make scientific research and industry more eco-friendly?

Updated laboratory equipment can play a significant role in increasing the sustainability of science. Manufacturers and equipment providers are turning to new methods to help reduce their carbon footprint. The latest technologies are enabling the development of reusable labware made from non-toxic materials, and sustainable manufacturing processes have led to more energy-efficient cold storage products, like ultra-low-temperature freezers.

Coupled with these technological advances, important initiatives such as the Accountability, Consistency and Transparency (ACT) program from My Green Lab empower laboratory managers to make better informed decisions when purchasing products. ACT and programs like it aim to help laboratories develop more environmentally conscious purchasing habits and adopt greener measures that will make the scientific sector more sustainable.

## The ACT Label

The ACT label contains several environmental impact scores for lab products, instruments, and reagents, much like an eco-nutrition label. It breaks down each product into several categories, including information about manufacturing (shipping impact and renewable energy use), user impact (water consumption and product lifetime), and end of life (how recyclable, compostable, or biodegradable the

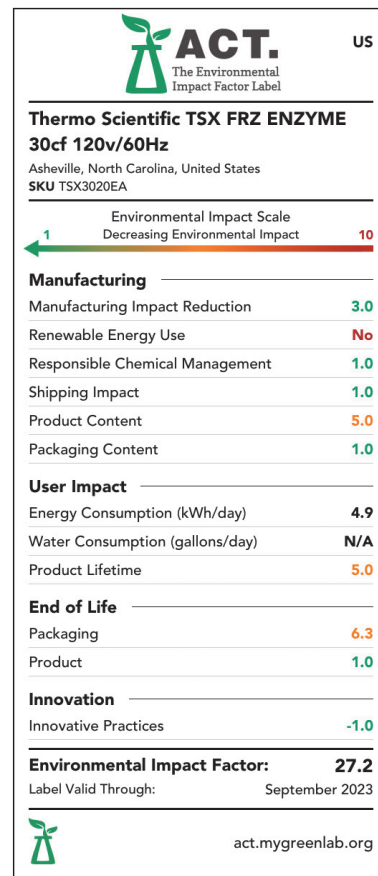
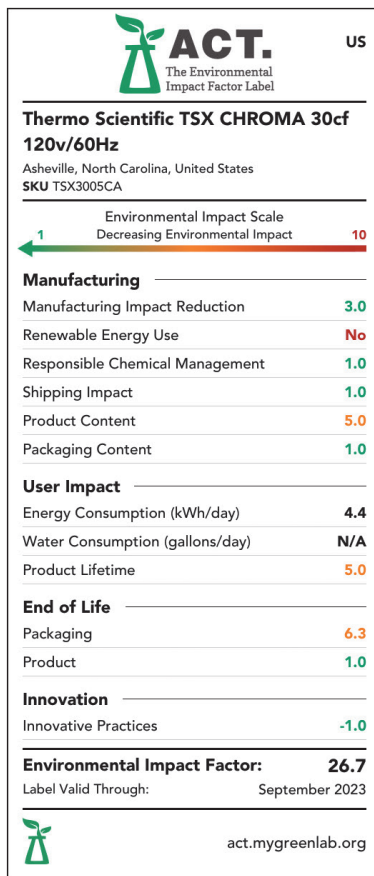
product is after disposal). By emphasizing the accountability (A), consistency (C), and transparency (T) of the environmental impact of laboratory products, it helps labs easily compare options and enables users to make more environmentally sustainable choices. The ACT label also gives manufacturers the opportunity to promote their commitment to sustainability by highlighting the areas in which their manufacturing processes are more efficient and sustainable.

While participation in the ACT program is not compulsory, it benefits the industry by encouraging manufacturers to improve the sustainability of their products, helping them understand how customers might value, for example, manufacturing using

renewable energy or more recyclable packaging materials. The hope is that this sets a precedent for the sector and helps customers make environmentally informed selections of hardware and services. The goal is to motivate companies across the equipment, reagent, and services supply chain to champion and participate in driving to make science more sustainable.

## Partners in Sustainability

Thermo Fisher Scientific has partnered with My Green Lab to highlight products that meet the ACT label criteria for sustainable manufacturing. Thermo Fisher's entire cold storage portfolio





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of more than 1,200 products will be ACT label-certified, enabling research, pharmaceutical, and clinical laboratories using these instruments to meet their sustainability objectives while preserving the integrity and viability of stored materials.

The first Thermo Fisher cold storage solutions to receive the ACT label were Thermo Scientific TSX Series Ultra-Low-Temperature (ULT) Freezers and Thermo Scientific Standard-Performance (STP) Ultra-Low-Temperature Freezers, with more than 200 models receiving certification. These state-of-the-art ULTs reduce energy usage up to 70 percent compared with conventional options. Thermo Fisher's ULT freezers were also among the first to receive ENERGY STAR certification from the U.S. Environmental

Protection Agency (EPA). The systems are powered by Thermo Fisher's unique V-drive technology, which detects and adapts to changing usage patterns and adjusts compressor speed as needed, offering significant energy savings without compromising performance.

In addition to ULTs, Thermo Scientific TSX Series High-Performance Laboratory Refrigerators and Freezers were the first cold storage units in the industry to receive ACT label certification. The new ACT label certification covers all models and sizes of TSX Series High-Performance Refrigerators and Freezers, including those designed for chromatography, blood banking, plasma storage, vaccine storage, and general-purpose laboratory use. TSX Series High-Performance Refrigerators and Freezers, which also use V-drive

technology, are ENERGY STAR certified and offer similar performance and energy saving capabilities.

## Our Commitment

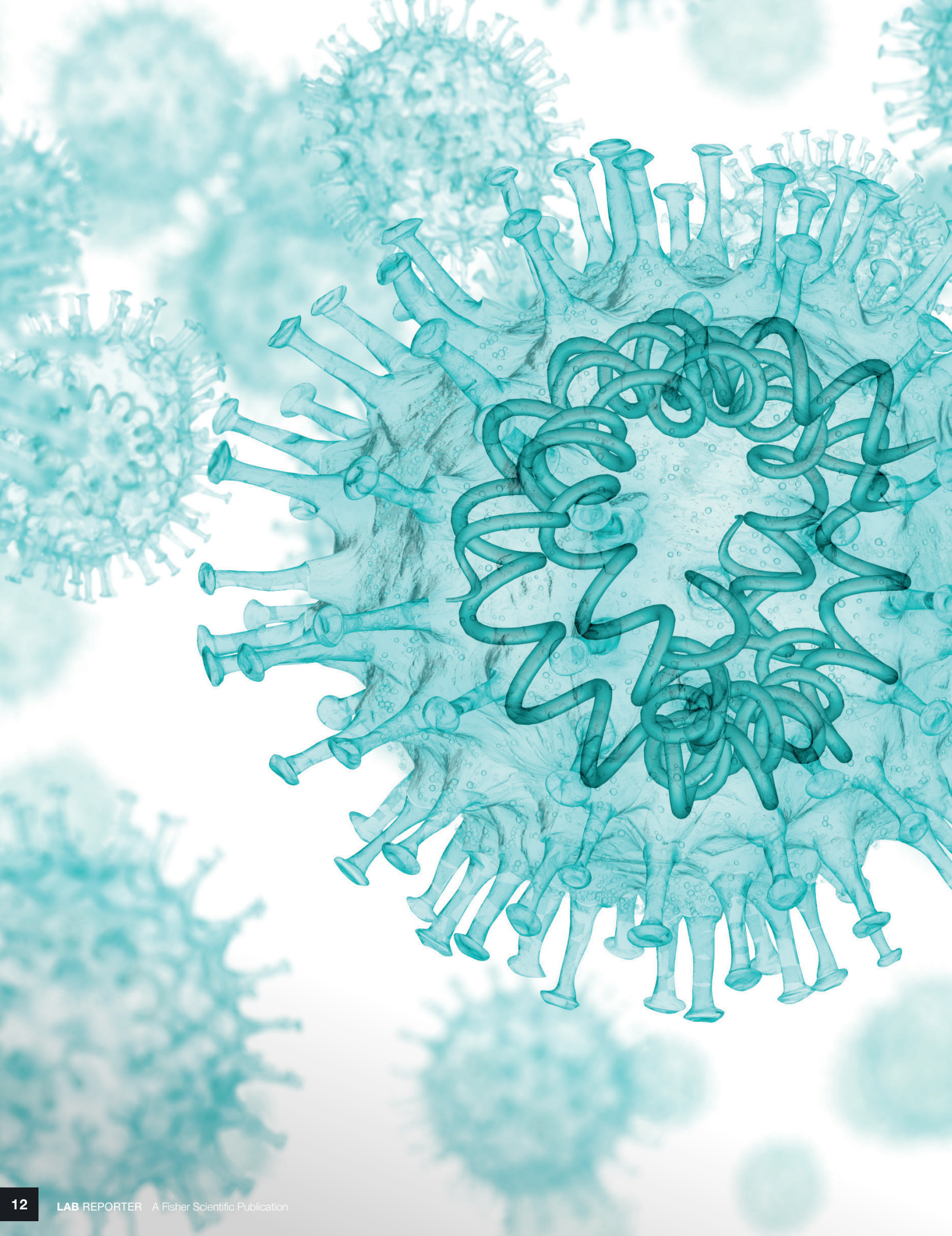
Thermo Fisher Scientific is committed to sustainable manufacturing for the entire TSX and STP product lines. In addition to using natural hydrocarbon refrigerants, Thermo Scientific cold storage products use SNAP-compliant, water-blown foam insulation that helps minimize heat emissions, allowing for considerably lower heating, ventilation, and air conditioning (HVAC) costs. The equipment is also manufactured in an award-winning, zero waste-to-landfill facility and care is taken to maintain eco-friendly supply chains, with raw materials purchased from sustainable sources. For example, TSX Series Refrigerators and Freezers are manufactured with increasingly long lifespans and shipped in packaging with high recycled and recyclable content and include recycling instructions for the recipient. At the end of a product's life, we can also help with repurposing and recycling.

New drives in the industry and a greater emphasis on sustainable manufacturing are helping scientists reduce the carbon footprint of their work. Moreover, initiatives like My Green Lab's ACT standard help customers make purchasing decisions based on a product's environmental impact and provide access to greener products and manufacturers. Together, manufacturing more eco-friendly equipment, greener supply chains, and environmentally conscious purchasing decisions can and must make a difference to the sector's record on carbon dioxide emissions.

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# Structure-Based Design: A Path Toward a Universal Coronavirus Vaccine

By Mark Miller

On January 3, 2020, the cause of several pneumonia-like cases in Wuhan City, China, was confirmed to be a novel *Betacoronavirus*. Three days later, Jason McLellan, PhD, a professor of molecular biosciences at the University of Texas, Barney Graham, PhD, an immunologist and virologist with the National Institutes of Health (NIH), and their colleagues agreed to determine the structure of the virus and help develop a vaccine. According to the article “Jason McLellan: the scientist stopping coronavirus in its tracks” written by Rebecca Pool, PhD and published in *Wiley Analytical Science*, the virus genome was sequenced and the researchers knew their next steps in just four days.

McLellan and his colleagues were in a position to respond so rapidly as the result of years of work developing structure-based vaccines to fight diseases like COVID-19. Here is a look at how they got there and what may be coming next.

## Locking the Protein

Around 2008, McLellan began working on the structures of HIV proteins. He then teamed up with Graham at the NIH Vaccine Research Center to apply a structure-based approach to the respiratory syncytial virus (RSV). By 2013, they had used X-ray crystallography to understand the structure of RSV’s fusion protein and how it changed states to infect cells. They were able to lock the protein in its prefusion state to prevent transmission and develop an initial vaccine.

“We were all so excited,” McLellan said. “We had used structural information to create a vaccine antigen that was superior to anything else ever created.” It was this breakthrough, according to Pool’s article, that laid the foundation for the work that would be done on the SARS-CoV-2 virus.

## 2P Mutation

McLellan departed NIH to run a lab at Dartmouth College. There, he and his team encountered the Middle East Respiratory Syndrome (MERS), a *Betacoronavirus* similar to SARS.

“The RSV and MERS fusion proteins are ancestrally related and have a similar fold in one part so we thought, ‘let’s give this a try,’” McLellan recalled. “I guess at the time of MERS we had been anticipating another coronavirus outbreak and so started

work on this quickly. Like RSV, we knew we wanted to stabilize that spike protein, make vaccine antigens, and isolate the antibodies.”

But there was a problem. The MERS spike proteins would not crystallize, which excluded X-ray crystallography studies. The team turned instead to cryo-electron microscopy with the help of specialist Andrew Ward from Scripps Research Institute.

Applying the new technique along with knowledge about the human coronavirus HKU1, McLellan and team became the first to resolve a human coronavirus spike protein with HKU1. From there, they stabilized the MERS spike protein by using proline to replace two of the protein’s residues, an amino acid substitution known as a 2P mutation.

## The Road Ahead

2P mutation proved a critical advancement when McLellan, now at the University of Texas, and the other researchers took on SARS-CoV-2.

“The SARS-CoV-2 was a *Betacoronavirus* and was very similar to the first SARS coronavirus, which was great in terms of rapidly developing a vaccine,” said McLellan. In a little over a month the team reported the results researchers needed to develop a vaccine based on the 2P mutation.

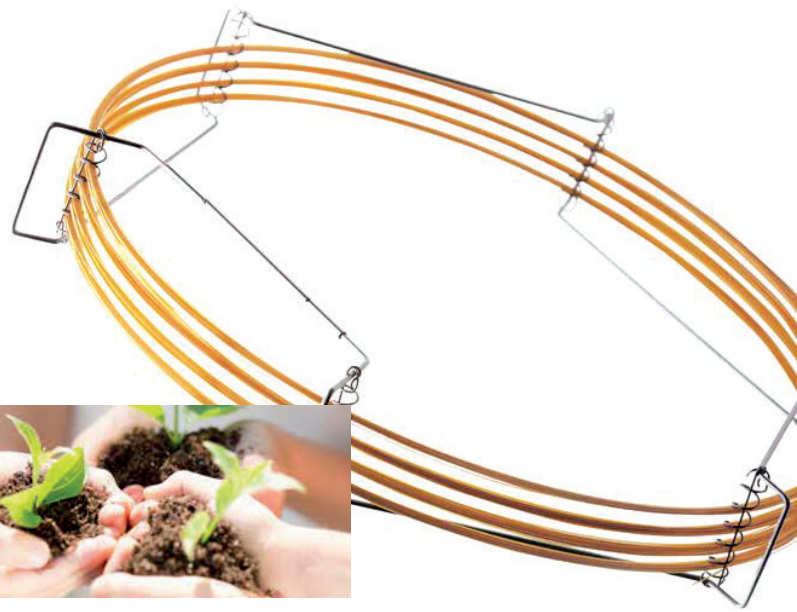
They had also discovered that proline could be added to the protein spikes of other coronaviruses, offering the potential for a universal vaccine — a single vaccine that could protect against many SARS-CoV-2 variants and emerging coronaviruses.

Since their initial work, four more proline molecules have been added to the structure to bring about a more stable spike protein known as a HexaPro. It’s in use in over 100 laboratories worldwide doing COVID-19 research, including vaccine development.

McLellan and his team continue to study coronaviruses and pursue a universal vaccine. In particular, they are investigating the virus’s S2 subunit, a strong target for vaccine development that’s shared across a range of coronaviruses.

*Mark Miller is a Thermo Fisher Scientific staff writer.*





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# BioClean Nitrile Isolator Gloves

## Greater Protection and Increased Longevity

BioClean Isolator Gloves are manufactured from nitrile with incredibly low levels of particles and excellent electrostatic discharge (ESD) properties. Designed for use in injectable manufacturing and pharmaceutical production areas, our clean or clean and sterile isolator gloves are cleanroom processed and packed and available to fit a range of port sizes.

### Advanced Protection and Dexterity for Workers in Pharmaceutical Manufacturing

Gloveboxes play a vital role in protecting products from human or environmental contamination as well as protecting individuals from workplace hazards. They're designed to provide a controlled, enclosed work environment that is separated from workers by a barrier, ensuring the containment of these sensitive and critical materials.

Every worker has the right to a safe workplace. When manufacturing specialized products in a pharmaceutical manufacturing environment, there are specific requirements for hand protection. Production and handling of these types of products typically happens in a clean or controlled environment and quite often in a sterile environment as well.

Due to the propensity of sensitive materials to be used in the life sciences, three types of gloveboxes may be used. Each is designed for the specific hazards present or level of cleanliness required.

Containment gloveboxes are designed to protect the operator and ambient environment from the material being processed. They are commonly found in pharmaceutical compounds relating to the development of oral treatments, neurological pain drugs, enhancing drugs, nuclear medicine, and other potent drugs.

Isolation gloveboxes protect the material being processed from the operator and the environment. Isolation gloveboxes are used in applications such as aseptic drug manufacturing and filling and parenteral (injectable) drug development.

Isolators combine features of containment and isolation gloveboxes and are used in the manufacture of cytotoxic parenteral drugs, some chemotherapy drugs, and biopharmaceutical cancer drugs.

Glovebox gloves provide a vital interface between the worker and the interior glovebox environment and must maintain a clean, reliable barrier while allowing the worker to effectively conduct manual tasks.<sup>1</sup> The gloves' integrity is crucial, as any breach in containment puts workers and products at risk of contamination.

### Materials

While these gloves have traditionally been manufactured with natural rubber latex and chorosulfonated polyethylene (CSM), recent advances in nitrile and other materials offer significantly improved protection, reliability, and longevity.

Nitrile is a synthetic, non-solvent based, FDA-compliant polymer with excellent anti-static properties that is ideal for use

with solvents and powders. Nitrile can withstand temperatures of up to 248°F (120°C) and is autoclavable. It can also be sterilized by gamma irradiation and sanitized with vaporized hydrogen peroxide (VHP) and isopropyl alcohol (IPA). Nitrile gloves can also be washed, processed, and packaged within a cleanroom environment, ensuring the gloves are an ultra-low contamination risk before being introduced into the isolator glove box.

Nitrile has superior puncture resistance, dexterity, and user comfort and offers



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excellent chemical resistance, providing extended permeation protection against many cytotoxins and the ability to maintain its properties even after gamma irradiation.

BioClean RABS and Isolator Gloves are manufactured from nitrile with incredibly low levels of particles and excellent ESD properties, offering safety to both the worker and the products being handled. The gloves are made with accelerator-free biodegradable nitrile and individually triple bagged in easy-open low-density polyethylene (LDPE) for convenient access.

BioClean gloves are 100% inspected and water leak tested and can be gamma irradiated, providing a Sterility Assurance Level of  $10^{-6}$ . They meet ASTM D6978-05 standards for handling chemotherapy drugs and offer an excellent resistance to VHP and IPA.

At Ansell, we are committed to delivering cutting-edge protection, enabling workers to increase quality and productivity without putting their health at risk. BioClean Nitrile Isolator Gloves are an excellent choice for improved comfort, protection, and performance.

1. American Glovebox Society  
AGS-G001-2007, <http://www.gloveboxsociety.org/index.asp>





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FEATURED ARTICLE

# Science Could Bring Sustainability to Your Table

By Kylie Wolfe



Meat is a staple in global diets. Whether you prefer chicken, beef, seafood, or no meat at all, you'll find it in most freezers, grocery stores, and lunch boxes. But with demand so high, the environmental consequences of meat production are causing concern.

That's why scientists have been looking for more sustainable ways to bring meat to the dinner table: some are turning to plant-based options and others to cultivated meats. The latter, an up-and-coming innovation, is a genuine meat product grown from real animal cells.

"While animals may have worked in the past as the chosen technology for producing meat, in the modern world it's really becoming a problem and it's only going to become more of a problem," said Claire Bomkamp, a senior scientist at the Good Food Institute, referring to issues ranging from climate change to antibiotic resistance. The goal is for alternative proteins to help restore balance. Instead of changing the way we eat and what we eat, scientists are working on viable stand-ins.

## Modernizing Meat Production

Environmental concerns regarding conventional meat are, in part, linked to greenhouse gas emissions. Livestock, for example, accounts for nearly 14.5 percent of these emissions globally.<sup>1</sup> Therefore, as population and demand continue to rise, sustainability concerns surface.

Resource use is another issue of traditional meat production and a piece of the climate change conversation. Cattle need large amounts of land to roam, plus ample food and water, to produce a finite assortment of meat. Chickens also need significant amounts of water to produce a single egg, 53 gallons, to be exact. To help, a company called Eat JUST is making plant-based eggs using 98 percent less water, 86 percent less land, and emitting 93 percent less carbon dioxide.<sup>2</sup>

"Meat is not going anywhere, so how can we make the same products that people love without the externalities that come with conventionally produced animal meat?" said Bomkamp. Nonprofits like the Good Food Institute and companies like Eat JUST are working on that answer, bringing science to the table.

Bomkamp explained that if cultivated meat is produced using sustainable energy sources, like solar and wind power, its environmental impact in terms of greenhouse gas emissions is likely similar to that of tofu or chicken. Even when produced using conventional energy, cultivated meat might be more of a friend to the environment than beef.<sup>3</sup>

## Creating Greener Alternatives

Cultivated meat production begins with a collection of animal cells. Scientists use these cells to establish cell lines that can be added to a nutrient-dense medium containing amino acids, vitamins, and other growth factors. This combination is ultimately placed in a bioreactor where, under the right conditions, the cells grow and multiply until they're harvested.<sup>4</sup> The resulting edible muscle and fat tissue can be made into hamburgers, sausages, and other meat products.

Bomkamp said this method is like plant cutting, taking a piece of one plant to grow a second. Even though this propagation technique differs from cell culture, it's a simplistic representation of what it means to cultivate meat. On a cellular level, cultivated and conventional meat are identical. With more research, taste, texture, and hopefully, consumer cost can be too.

"Meat is not going anywhere, so how can we make the same products that people love without the externalities that come with conventionally produced animal meat?"

Depending on the type of meat a company produces, the overall process, from cell sample to finished product, takes anywhere from two to eight weeks. It's thought to be more efficient and environmentally friendly than agriculture in the traditional sense, but more studies are needed to determine the extent. Scientists also believe that cultivated meat could help lower cases of foodborne illness because it will be raised in sterile conditions where contamination risk is kept to a minimum.

In March 2019, the Food and Drug Administration (FDA) and United States Department of Agriculture (USDA) agreed to oversee certain steps of the cultivated meat production process.







continued from page 21

# Science Could Bring Sustainability to Your Table

For livestock and poultry, the FDA oversees cell collection, growth, and differentiation, and the USDA oversees harvesting, processing, packaging, and labeling. For other meat products, the FDA manages each step.<sup>5</sup> Their joint goal is to implement food safety regulations and provide accurate labeling for the public.

As of December 2020, the Singapore Food Agency approved a cell-cultured chicken product that became the world's first cultivated meat approved for sale.<sup>6</sup>

## Gauging Sustainability and Success

Bomkamp says that the short-term success of this industry means giving those who love meat, but don't love its environmental consequences, access to an alternative. This also means creating a product with the same taste and texture as traditional meat — with a smaller environmental footprint. As we enter the future of meat production, scientists want to find a balance between what's good for people and the planet. But that may take time.

“Do we get to the point where you can go to the grocery store and buy these products? That's certainly the hope,” said Bomkamp. “But predicting timelines from the beginning is a little bit of a crystal ball situation.”

Aside from solidifying the science and scaling production to generate tasty, environmentally friendly, and cost-effective meats, public acceptance may also hinder the reach of alternative proteins and the time it takes for them to become mainstream.

According to a study published in *Frontiers*, 29.8 percent of survey participants said they were likely to purchase cell-cultured meat. The remaining 70.2 percent of participants were either unlikely or only somewhat likely to do so.<sup>7</sup> Greater transparency from companies and government agencies can help raise awareness for cultivated meat, but it's up to consumers to decide whether alternative proteins are welcome or unwelcome on their plates.

## Deciding the Future of Meat

It's possible to reallocate resources and potentially curb the meat industry's contributions to climate change, but the path forward is to be determined. Scientists, legislators, and regulators are hoping to find an approach that's better for the environment and meets public demand, even if it means changing the way we define our proteins.

“What if we not only switched to cultivated meat but were also really intentional with the land that's freed up and no longer used for the production of food? What if we used that land for rewilding or other climate-positive activities?” Bomkamp said. “We should really be thinking about how to take advantage of those sorts of synergies.”

More research is needed to understand the environmental significance of cultivated meat production. Studies about water, energy, and land use will help tell a clearer story about alternative proteins and their role in the future of the food industry.

*Kylie Wolfe is a Thermo Fisher Scientific staff writer.*

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# Optimize Cell Growth with a Biological Shaker

For decades, shaking has been used to cultivate a variety of cells, including bacteria, fungi, and plant and animal cells in suspension. This agitation helps to increase the availability of nutrients and improve oxygen transfer, ultimately providing higher cell yields than static incubation.

Biological shakers help to automate and standardize this task. They're an easy-to-use and inexpensive choice for basic applications like organism screening, media design, inoculum preparation, and early process development. It's hard to imagine a biotechnology lab that doesn't use shakers, but how can you use them to maximum benefit?

Here are a few things to consider when using a shaker to grow cells.

## Agitation Modes

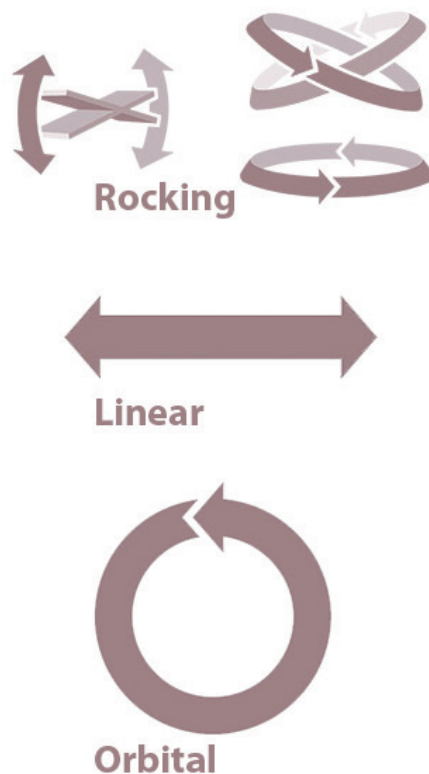
There are three different types of agitation to choose from: rocking, linear shaking, and orbital shaking. While these motions differ, they're similarly achieved: liquid in a vessel is placed on a platform that moves in a particular manner, which agitates the liquid.

Rocking shakers move in either a two- or three-dimensional see-saw motion about a central point. This motion creates a wave-like mixing process that's generally used for gentle mixing and incubation procedures, including DNA extraction, staining and de-staining, and washing blots and gels. For cell cultivation, you can even find specifically constructed rocking devices that allow mammalian cells to be cultivated in single-use bags.

Linear shakers, also known as reciprocating shakers, move the liquid back and forth in a single plane. These

shakers have largely vanished from labs cultivating cells because of their primary drawback: While speeding up, linear shakers can cause unpredictable geyser-like eruptions that can wet the flask's closure, potentially contaminating the sample and decreasing aeration.

Orbital shakers are the most common choice for cell expansion. These shakers rotate the platform in a circular motion, creating a consistent "swirl" pattern. They're available with a variety of options



for specific culturing demands, such as temperature control, photosynthetic lighting, and CO<sub>2</sub> control.

## Shaking Speed

Agitation not only improves oxygen transfer but also maintains homogeneous conditions in the medium through continuous mixing. In general, higher agitation speeds (as well as larger orbits) increase surface area for gas exchange, providing better aeration and higher yields.

However, higher speeds also lead to shear forces that can negatively influence microorganisms, potentially causing changes in morphology, variations in growth, and even cell death. While bacterial and yeast cells can withstand and even benefit from higher speeds, mammalian cells are more sensitive and require lower agitation speeds.

## Agitation Orbit

Orbital shakers have common diameters of 19, 25, and 50 mm. As a rule of thumb, 19- and 25-mm orbits are still the standard in microbiology.

Larger orbits can increase the surface area of samples at speeds of 150 rpm and slower, so they're useful when working with cells that are sensitive to shear forces. Larger orbits are also the best choice when working with vessels larger than two liters and wide vessels like Fernbach flasks.

Agitation Speed		100 rpm	200 rpm	300 rpm
OTR (mmol O <sub>2</sub> /L/h)	25 mm orbit	10.3 ± 0.2	62.0 ± 1.1	186.1 ± 0.5
	50 mm orbit	12.0 ± 0.4	86.8 ± 7.8	214.0 ± 5.2

OTR measurements in a shaker with different set speeds in a 25 vs. 50 mm orbit shaker. Results shown were the average of duplicated experiments (Internal testing, Eppendorf, 2017)



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But larger orbits and higher speeds don't always lead to higher growth rates. Flask design, flask material, and filling volume are also important considerations.

## Flask Design

Introduced at the beginning of the last century, shaker flasks have a variety of benefits. They're available in a wide range of sizes from 25 mL to 5 L, making them applicable for a variety of experiments from screening and expansion to media design and early process development. And they're affordable and suited to cultivation of bacteria, yeast, fungi, and plant and animal cells in suspension.

Typical Erlenmeyer flasks feature conical bodies, wider bases, and cylindrical necks. However, specialized designs can further improve gas exchange: Fernbach flasks offer wider bases that provide a large area for oxygen transfer, and Eppendorf Ultra

Yield and Optimum Growth flasks feature optimized shapes that maximize surface-to-volume ratio.

## Glass vs. Plastic

Both glass and plastic flasks have their benefits. For most classic microbiology applications, autoclavable glass Erlenmeyer flasks are a durable and reusable choice. But when contamination matters — as in production or handling sensitive cultures — sterile, single-use disposable flasks with 0.2 µm filter vented caps offer the most convenience and safety.

## Filling Volumes

Lower filling volumes are best for cultures that have a greater need for aeration. By using a smaller percentage of a vessel's total volume, you can create a larger surface area for gas exchange, which leads

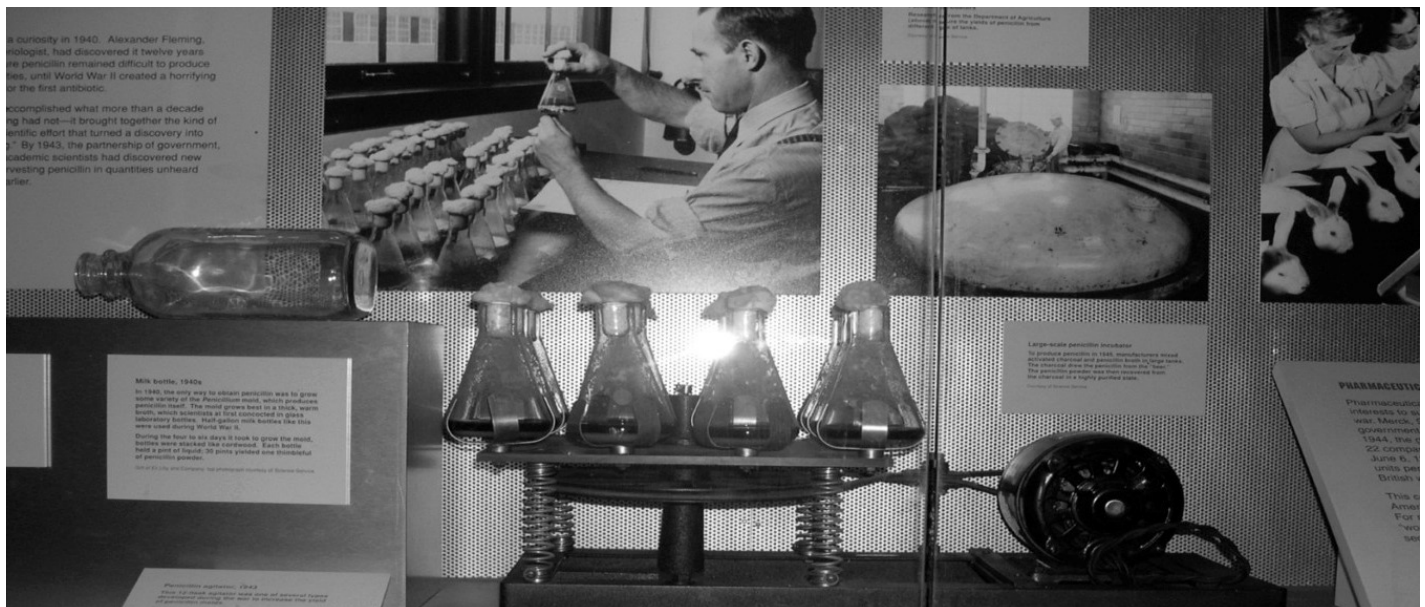
to higher cell yields.

As a rule of thumb, don't exceed 20 percent of nominal flask capacity for microbial cultures. For maximum oxygen transfer, reduce filling volume to as little as 10 percent and increase rotational speed as much as your cells can withstand before succumbing to shear forces.

For mammalian cultures sensitive to shear forces, choose a fill volume between 30 and 40 percent and a shaking speed no higher than 150 rpm.

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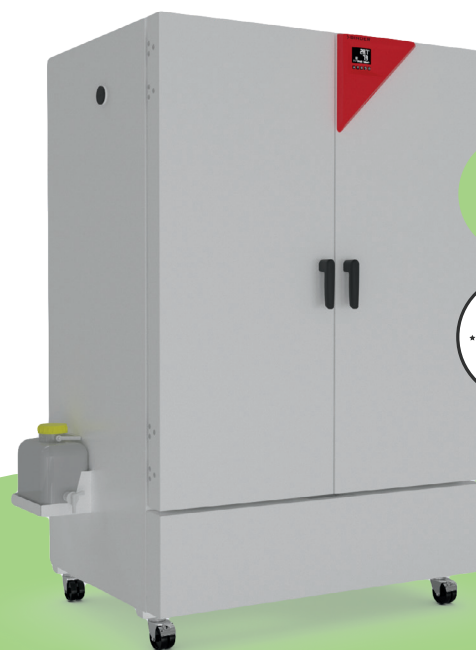
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# Wash Away Worry with an Automated Glassware Washer

By Jordan Henderson, Product Manager, Labconco

Today's laboratories require a steady supply of clean labware to operate efficiently. While cleaning glassware, plasticware, and other utensils is a relatively mundane task, it can have an enormous impact on the quality, accuracy, and precision of your lab's results.

Washing glassware by hand is common in many laboratories, but it inherently varies from person to person, which can cost your lab time, energy, and money if it negatively impacts your results. Automated glassware washers, on the other hand, can help you more consistently meet cleanliness goals for your labware.

Automated washers come with a variety of features, so find one that offers the efficiency, versatility, and reliability you need.

## Efficiency

Glassware, plasticware, and utensils often require different methods of washing and rinsing. Whether your labware needs robust cleaning from consistent, efficient spray arms or more targeted cleaning from direct injection washing and rinsing with spindles, your glassware washer should be able to easily accommodate whatever approach best suits your laboratory.

Your washer should also provide sufficient water temperature to adequately clean your labware. Some glassware washers may heat water to a maximum temperature of only 70°C (158°F) and lack the ability to tackle the boiled, baked, and caked-on contaminants many labs see every day. Wash and rinse water temperature minimums of 93°C

(199°F) soften and penetrate stubborn contaminants while providing thermal disinfection, further reducing potential carryover from one wash cycle to the next.

It's also important to find a washer that can keep up with high demand without sacrificing performance or cleaning ability. Choose a washer that's intentionally designed to do more in less time with fewer resources. For example, the time it takes a washer to heat wash and rinse water directly affects the length of wash programs. A washer that allows you to choose between single-phase and three-phase heating options can provide more robust heating for the shortest, most efficient wash cycles.

Because automated glassware washers are far more consistent than hand washing, they can also help you standardize consumption of resources like water and electricity — they can even help you save time and money. Implementing an automatic glassware washer will consistently provide an excellent level of

glassware cleanliness from the first cycle to the last, helping you avoid the negative impact that dirty labware can have on results.

## Versatility

Rack flexibility is key when assessing your glassware washer. Glassware and plasticware come in all shapes and sizes, and your washer racks should be flexible enough to tackle whatever combination of labware you throw at them. When selecting a washer, look for useful features like adjustable rack positions, removable components, and overall functionality with available inserts, like those that hold test tubes or trays.

Need to wash vials, pipettes, or other pieces of specialized glassware? Glassware washer accessories should complement not only the washer but also your atypical, difficult-to-clean flasks, pipettes, and other specialty glassware.

The user interface is also an important consideration. It should deliver an

Description	Cat. No.
SteamScrubber, 115V, 60Hz, 1-Phase	40-100-1000
SteamScrubber, 208/230V, 50/60Hz, 1-Phase	40-100-1010
SteamScrubber, 208/230V, 50/60Hz, 3-Phase	10-100-1013
FlaskScrubber, 115V, 50/60Hz, 1-Phase	41-100-1000
FlaskScrubber, 208/230V, 50/60Hz, 1-Phase	41-100-1010
FlaskScrubber, 208/230V, 50/60Hz, 3-Phase	41-100-1013
FlaskScrubber Vantage, 208/230V, 50/60Hz, 1-Phase	42-210-1010
FlaskScrubber Vantage, 208/230V, 50/60Hz, 3-Phase	42-210-1013

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intuitive experience that allows a user to tailor wash programs for specific workflows. Some washers even offer onboard data logging, storage, and export via USB or a secured network connection, which can help you validate custom wash programs.

## Reliability

Water quality can vary from region to region, so using pure-water rinses is a great way to further ensure that your labware is clean. Water qualities such as hardness and pH level, as well as the presence of silica, metals, chloride, or organic materials, can result in residues that have a significant impact

on the quality of your results. However, pure-water pumps aren't standard on all automated glassware washers. Be sure to check for this — it can save you the headache of upgrading your current system or installing a new water purification system if the need for pure water rinses arises in the future.

The longevity and durability of a washer depends on the quality of its internal components. Underpowered components result in underpowered washers. When selecting a washer, be sure it uses robust pumps and quality valves and internal assemblies to minimize unscheduled down times.

In the unlikely event of mechanical issues, you should be able to quickly assess and identify component failures. A standard array of diagnostic tests can help identify potential component failures, and a warranty and post-installation support can help you quickly complete repairs. Similarly, support from knowledgeable service technicians can have a huge impact on getting your washer back up and running.

When considering an automated glassware washer, efficiency, versatility, and reliability should be standard on the model you choose. By focusing on these aspects, you can free scientists from the sink and help them get back to the lab.







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# Who to Hire?

## Lab Staff Can Affect Publication Output

By Mike Howie

When asked to do more with less, how can you optimize productivity? How can you complete not just a large amount of work but effective, successful work with limited time, money, and resources? While these questions have been buzzing in many minds recently, they're not new.

Back in 2015, a pair of researchers — Annamaria Conti of the Georgia Institute of Technology and Christopher Liu of the University of Toronto — investigated how the composition of laboratory staff can affect publication output. They found that simply hiring more people doesn't necessarily help you get more done.

### The Study

Conti and Liu focused their research on the biology department at the Massachusetts Institute of Technology, examining lab staffs and output from 1966 to 2000. Their dataset included 119 principal investigators and 5,694 laboratory members, who in total published 7,844 papers. About 15 percent of those papers were published in *Science*, *Nature*, or *Cell*.

The average laboratory in the study included about five postdocs, three graduate students, and two technicians. Using regression analysis, the researchers found that adding one member to a lab of this size correlated with an extra quarter publication. While adding even more members to the lab further increased productivity, it did so at slower and slower rates until lab size reached 25 people. After that, adding more staff decreased productivity.

What's perhaps more illuminating, however, is the fact that some employees seemed to have a greater impact than others. For example, adding a graduate student correlated to an extra 0.14 publications while adding a postdoc correlated to an extra 0.31. And even among postdocs, those with grant support were linked to 0.19 additional publications while those with fellowships were linked to an additional 0.29. Adding technicians, it seems, didn't make a difference.

The story is slightly different when only considering work published in *Science*, *Nature*, or *Cell*. While bigger labs still seemed to publish more often, the researchers found that adding a lab member of any position increased the likelihood of publication by eight percent. Once lab size reached 22 people, however, hiring more became counterproductive.

Position within the lab still had some effect on publishing in these high-impact journals, but it differed from other publications. This time, graduate students seemed to have increased the likelihood of publication just as much as postdocs, as did technicians. But postdocs without fellowships did not increase the probability of publication.

The study, *Bringing the lab back in: Personnel composition and scientific output at the MIT Department of Biology*, was published on *ScienceDirect*.

### Remaining Questions

While the study found thought-provoking correlations, it didn't identify any specific causality. It didn't determine why, for example, postdocs with fellowships had more impact than those without fellowships, or why graduate students seemed more beneficial than technicians. Liu speculates that graduate students may be better positioned to work on long, risky projects.

"People at different training stages make different contributions," Liu explained, "but it's not that they're less productive. They just have different productivity in different projects."

There could be a variety of reasons why one employee has more impact on publication than another, and it's possible that the study's conclusions may not hold within other schools or disciplines. What remains consistent is that the people we hire are critical to our success.

*Mike Howie is a Thermo Fisher Scientific staff writer.*



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Model	Capacity	Cat. No.
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FB-11203	5.75 L (1.5 gal.)	<b>FB11203</b>
FB-11206	6.9 L (1.8 gal.)	<b>FB11205</b>
FB-11207	12.75 L (3.3 gal.)	<b>FB11207</b>
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ST 300 H	30 L	<b>08-000-100</b>
ST 500 H	49 L	<b>08-000-101</b>
ST 600 H	57 L	<b>08-000-102</b>
ST 800 H	83 L	<b>08-000-103</b>
ST 1400 H	126 L	<b>08-000-104</b>
ST 1600 H	162 L	<b>08-000-105</b>
ST 2500 H	255 L	<b>08-000-106</b>





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M	1,000/Case	9102	19-423-102
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Description	Quantity	Cat. No.
AquaPur	4 x 1 gal.	04-355-124
AquaPur	5 gal.	04-355-125
AquaPur	55 gal.	04-355-126
AquaPur ST	4 x 1 gal.	22-281-500



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Single Stage	20 L/min.	75 torr	15 psig	1/6 hp	13 x 6.5 x 8.5 in.	<b>13-880-14</b>
Single Stage	40 L/min.	65 torr	15 psig	1/6 hp	13 x 6.5 x 8.5 in.	<b>13-880-16</b>
Two Stage	20 L/min.	6 torr	15 psig	1/6 hp	15 x 6.5 x 8.5 in.	<b>13-880-18</b>
Two Stage	35 L/min.	6 torr	15 psig	1/8 hp	16 x 7 x 9 in.	<b>13-880-20</b>
Two Stage	35 L/min.	1.5 torr	15 psig	1/8 hp	16 x 7 x 9 in.	<b>13-880-22</b>

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Model	Application	Ultimate Vacuum	Flow Rate	ATEX Compliant	Integral Gas Ballast Valves	Cat. No.
N 96	Filtration, SPE, Aspiration	97.5 torr, 130 mbar	7 L/min.	No	No	<b>13-880-904</b>
N 820 G	Rotary Evaporation, Degassing, Fluid Aspiration, Centrifugal Concentration, Vacuum Oven, Gel Drying	4.5 torr, 6 mbar	20 L/min.	Yes	Yes	<b>13-880-905</b>
N 840 G	Rotary Evaporation, Filtration, Centrifugal Concentration, Vacuum Oven	4.5 torr, 6 mbar	34 L/min.	Yes	Yes	<b>13-880-906</b>

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Model	Applications	Capacity	Power	Cat. No.
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120	<ul style="list-style-type: none"> <li>Cell Disruption</li> <li>Protein Extraction</li> <li>DNA Shearing/ChIP</li> </ul>	0.2 to 50 mL	120 w	<b>FB120110</b>
505	<ul style="list-style-type: none"> <li>Cell Disruption</li> <li>Nanoparticle Dispersion</li> <li>Homogenization/Mixing</li> </ul>	0.2 to 1000 mL	500 w	<b>FB505110</b>
705	<ul style="list-style-type: none"> <li>Cell Disruption</li> <li>Protein Extraction</li> <li>DNA Shearing/ChIP</li> <li>Nanoparticle Dispersion</li> <li>Homogenization/Mixing</li> <li>Sonochemistry</li> </ul>	0.2 to 1000 mL	700 w	<b>FB705110</b>



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